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Guillaume Gines, A Zadorin, J.-C S Galas, T S Fujii, A S Estevez-Torres, et al.. Microscopic agents programmed by DNA circuits. *Nature Nanotechnology*, 2017, 12 (4), pp.351-359. 10.1038/nnano.2016.299 . hal-01489464v1

HAL Id: hal-01489464

<https://espci.hal.science/hal-01489464v1>

Submitted on 28 Apr 2017 (v1), last revised 28 Dec 2017 (v2)

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Microscopic agents programmed by DNA circuits.

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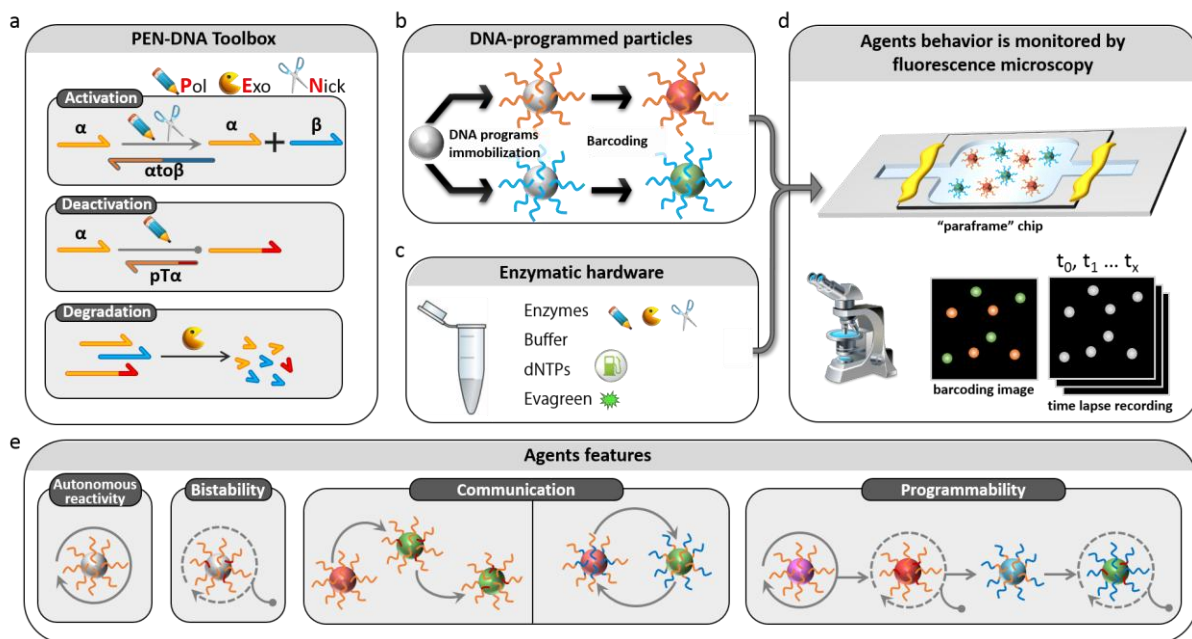
Abstract

Information stored in the sequence of synthetic nucleic acids can be used *in vitro* to create complex reaction networks implementing precisely programmed chemical dynamics. We report here the extension of this approach to program the local and individual chemical behavior of microscopic “agents” dispersed in an enzymatic solution. Agents possess multiple stable states, thus maintaining a memory, and communicate by emitting various orthogonal signals and sensing the behavior of neighboring agents. We build on these elements to create collective behaviors involving thousands of different agents, for example retrieving information over long distances, or creating spatial patterns. This approach recapitulates some fundamental mechanisms of distributed decision making and morphogenesis among living organisms. The possibility to scale up the information-processing capability of DNA-encoded artificial systems could also find applications where many individual clues need to be combined to reach a decision, e.g. in molecular diagnostics.

27 Molecular programming (MP) holds that information can be processed using molecules as carriers
 28 and chemical reactions as computational primitives. In the last decade, this emergent field has
 29 crafted an array of molecular mixtures that indeed behave as *systems*, in the engineering sense,
 30 even if they are essentially amorphous chemical soups^{1,2}. Many of these man-made systems are
 31 inspired by existing information-processing molecular networks observed within single cells: logic
 32 circuits seen in signaling cascades^{3,4}; circadian oscillators pacing the metabolism of light-harvesting
 33 microorganisms^{5,6}; all-or-nothing switches embedded in genetic regulation pathways^{4,7,8}, shape-
 34 forming morphogen fronts in developing embryos⁹⁻¹², etc.

35 In many cases, however, biological behaviors result from the collective dynamics of many cells
 36 exchanging molecular information. Bacteria use quorum sensing to collectively select survival
 37 strategies^{13,14}, multiple cells within embryos exchange morphogens to generate sharp differentiated
 38 clusters^{15,16}, and unicellular organisms such as *Dictyostelium* switch to a multicellular lifestyle to
 39 overcome adverse conditions¹⁷.

40 In this work, we introduce an experimental approach combining existing solution-phase, well-mixed
 41 MP techniques and solid-phase DNA biochemistry to create programmable individual microscopic
 42 objects. Each particle carries a stable set of DNA instructions controlling its dynamical behavior. It is
 43 also able to interact with its neighbors using diffusible signals, providing a way to design collective
 44 behaviors involving thousands of -possibly different- agents.



45
46

47 **Figure 1. Programming microscopic agents using DNA-encoded reaction rules.** *a*, The PEN DNA toolbox is a
 48 set of generic modules that can be cascaded to build molecular programs processed by an enzymatic
 49 machinery. *b*, In this study, we convert the solution-phase system to a spatially discrete architecture by
 50 attaching the rule-encoding templates to micrometric particles. *Particles* are barcoded to be easily
 51 distinguished by fluorescence according to the set of templates they carry. *c*, The reaction mix contains the
 52 enzymatic hardware in addition to the buffer, the fuel dNTPs and a double strand-specific dye EvaGreen. *d*,
 53 Particles are incubated in a 2-dimensionnal micro-chamber and the reaction (creation of output) is monitored
 54 by microscopic imaging of the *EvaGreen* fluorescence signals, reflecting the relative amount of active (i.e.
 55 double-stranded) templates on each particle. In the case of mixed populations, barcode channels are used to
 56 identify the circuit carried by each particle. *e*, Programmed particles behave as *independent* microscopic agents
 57 displaying features such as bistability, communication and programmability.

58
59

60 We build on the Polymerase/Exonuclease/Nickase Dynamic Network Assembly (PEN DNA) toolbox^{5,18},
61 a molecular programming scheme where short oligodeoxyribonucleotides (templates) encode the
62 connectivity information of a network of activating/inhibiting interactions (Fig. 1a). We attach these
63 rule-encoding DNA templates to micrometric porous particles (Fig. 1b). The particles are then
64 immersed within a solution containing the PEN enzymes, leading to the interpretation of the
65 particle-bound DNA rule into a local chemical behavior (Fig. 1c-d). We show that the individual
66 behavior of a bead carrying a tethered PEN DNA network qualitatively reproduces the one observed
67 for solution-phase systems with the same set of templates. However, production of DNA signals is
68 now spatially constrained to the position of the bead, while degradation extends to the bulk of the
69 solution, resulting in the generation of concentration gradients around each agent (Supplementary
70 Notes 1). This creates autonomous, mono- or multi-stable particles that communicate locally
71 through diffusive exchange of signals (Fig. 1e).

72 These building blocks enable the rational molecular programming of population-scale
73 phenomena^{19,20}. We generate spatially random arrays containing thousands of particles in which we
74 observed the propagation of bistable travelling fronts, or the generation of stationary symbiotic
75 activation patterns, depending on the molecular instructions carried by the microscopic agents. We
76 also show that multiple orthogonal layers of chemical communication can be used in a single system.
77 We used this capability to build an assembly where discrete agents cooperate to propagate chemical
78 information over a distance of a few millimeters, check for the presence of a particular bead type,
79 and carry this information back to the origin of the query (Fig. 1e, right).

80 **Particles encoded with a positive feed-back loop (PFL) program**

81 The dynamics of PEN DNA systems, relies on a generalist enzymatic machinery that drives the
82 exchange of DNA signals between the rule templates: a DNA-polymerase elongates an input strand
83 that hybridizes on the input (3') side of a matching template; a nickase recognizes and cuts the
84 resulting full duplex, releasing both the input and a new output DNA. The exonuclease non-
85 specifically degrades all unprotected oligonucleotides (templates are protected), maintaining the
86 system in a responsive out-of-equilibrium state. In functional terms, hybridization and enzymatic
87 reactions collaborate to generate a set of basic modules that can be cascaded in dynamic circuits
88 whose parameters are linked to kinetic, thermodynamic and concentration values. This versatile
89 system has already allowed the construction of various solution-phase circuits such as oscillators⁵ or
90 multistable switches⁸.

91 We investigated the possibility of grafting rule-encoding PEN DNA templates onto solid supports in
92 order to create spatially localized DNA programs^{21,22} (see Supplementary Information Methods
93 section, all sequences are given in Supplementary Table 1). **At first, streptavidin-conjugated porous
94 microparticles ($\varnothing = 34 \pm 10 \mu\text{m}$) were conjugated to a biotin-modified template αox , a dual-repeat
95 sequence catalyzing the positive feedback loop (PFL) reaction $\alpha \rightarrow 2\alpha$ (Fig 2a). Details concerning the
96 properties of the conjugated particles are given in Figures S1-4. These PFL-encoded particles ($\text{P}\alpha_{\text{M}}$)
97 were transferred to a master mix containing PEN enzymes, dNTPs and incubated in a flat microscopy
98 chamber. **The amplification reaction is monitored using a double strand specific dye (EvaGreen) that
99 fluoresces upon binding the active templates producing the α strand.** Time lapse fluorescence
100 imaging revealed the expected behavior for the PFL circuit: an initial exponential amplification phase
101 is followed by a steady-state plateau (production balances diffusion and degradation), then a return
102 to the baseline after exhaustion of dNTPs (Fig. 2c and Supplementary Movie M1).**

103 **Diffusion cone + theory...**

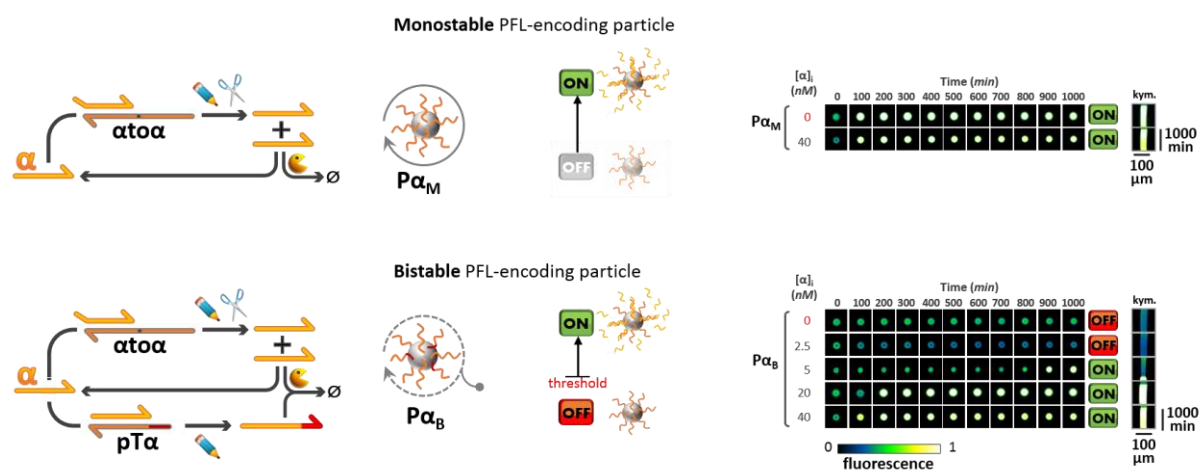
104

105 We first validated biotin modifications as a tethering strategy (Fig S1-2). We grafted femtomoles
106 (billions of copies) of biotinylated template oligonucleotides to streptavidin-modified porous beads

107 with a diameter of $34 \pm 10 \mu\text{m}$. When the particles were grafted below their binding capacity, we
 108 observed a shell-like geometry and negligible exchange of strands between the beads (Fig. S3-4). We
 109 then used standard PEN DNA rules^{5,18} to create a biotin-modified template $\alpha\text{to}\alpha$, a dual-repeat
 110 sequence catalyzing the Positive Feedback Loop (PFL) reaction $\alpha \rightarrow 2\alpha$ (Fig 2a). Particles encoded
 111 with this strand were transferred to a master mix containing PEN enzymes and dNTPs and incubated
 112 in a flat microscopy chamber. Time lapse fluorescence imaging revealed the expected behavior for
 113 the PFL circuit: an initial exponential amplification phase is followed by a steady-state plateau
 114 (production balances diffusion and degradation), then a return to the baseline after exhaustion of
 115 dNTPs (Fig. 2c and Supplementary Movie M1). This behavior closely parallels the one observed for
 116 the case of freely diffusing templates (Fig. 2b). In the case of particle-tethered templates however,
 117 change in fluorescence signal is observed only on the beads and not in the bulk of the solution,
 118 showing that production is indeed localized. The particles also acted in synchrony, suggesting that
 119 diffusion of signal strands and/or fuel can mediate communication.

120 The dimensionless Damköhler number Da gives an estimation of the importance of local versus
 121 global effects in chemical systems. Here the Da associated to grafted particles is much greater than
 122 1 (details of the estimation of Da and reaction-diffusion length scales are given in Supplementary
 123 Note 2), so we expect a dominance of local effects. Ultimately, a single particle **performing an**
 124 **autocatalytic reaction** should be able to **conserve** alone (on its own) a high production of output,
 125 despite being constantly depleted by diffusion. To check **this self-sustained reactivity**, a unique
 126 particle was incubated in a large chamber (volume ratio of 10^{-6} , Fig. 2d). Here, the total amount of
 127 template would be too small to sustain amplification in a bulk solution ($\sim 3 \text{ fmol}$ for a total volume of
 128 $10 \mu\text{L}$, i.e. less than **one** nanomolar). The strands are however highly concentrated within the
 129 particle, and we did observe a sharp localized amplification. The following plateau in fluorescence
 130 level was sustained for more than 18 hours, indicating that dNTP consumption by the single particle
 131 was limited. Amplification and homeostasis are therefore individual properties of particles bearing
 132 **autocatalytic** circuits, and not a global feature of the heterogeneous mixture.

cone de diffusion



133

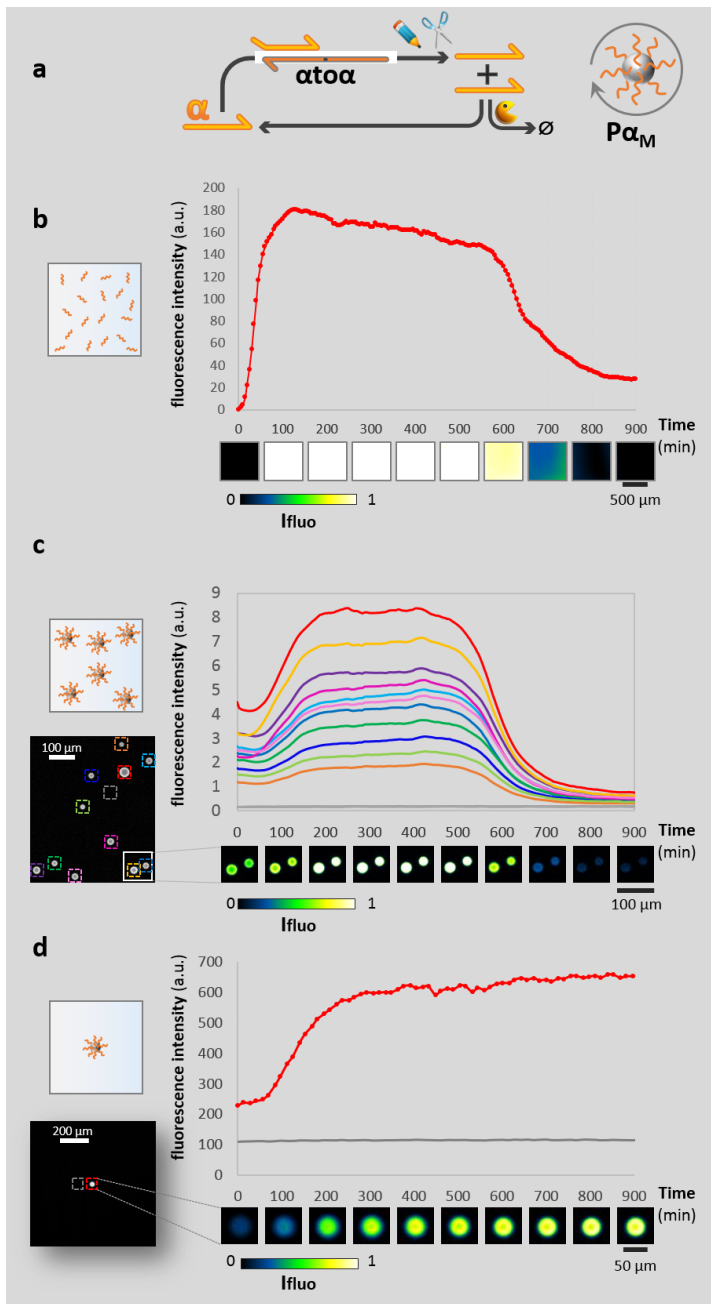


Figure 2. Agents carrying positive feedback templates are autonomous. *a*, A PFL encoded by template $\alpha\text{to}\alpha$ catalyzes the self-activated production of strand α . *b*, *c*, *d*, Microscopic fluorescence recording of the reaction in a flat micro-chamber *in absence of initial trigger α* for: *b*, freely diffusing templates (conjugated to streptavidin); *c*, Multiple microscopic porous particles functionalized with the template $\alpha\text{to}\alpha$; *d*, A single particle (particle to chamber volume ratio is $1 : 10^6$). *The amplification reaction is monitored with a double strand specific dye that fluoresce upon binding to the –double-stranded-active template.*

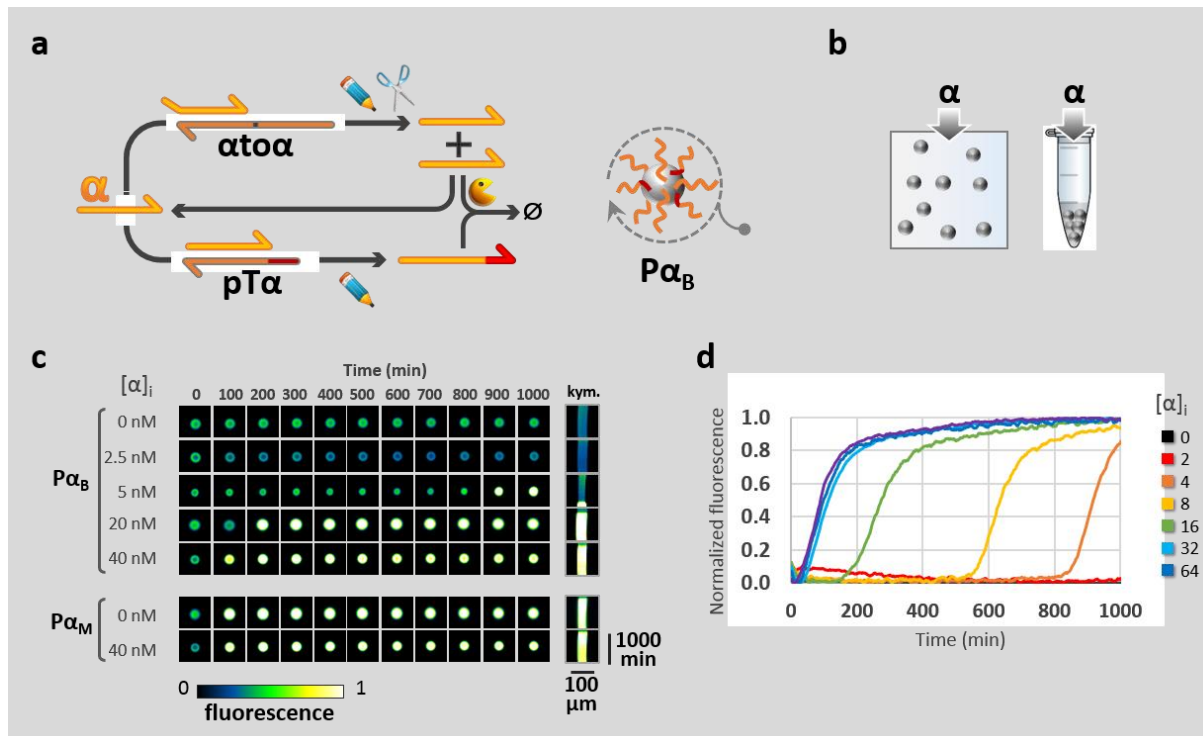
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135 Bistable agents

136 In the cases described so far we observed that, even in the absence of initial trigger, particles
 137 carrying sufficient $\alpha\text{to}\alpha$ inevitably self-activate after a short period of incubation. This phenomenon
 138 is again consistent with the behavior reported for solution-phase amplification systems:
 139 autocatalytic templates self-start after some time²³. This is inherent to the monostability of the first-
 140 order PFL network enforced by the dual repeat template: any small perturbation, such as leaky
 141 (primer-independent) polymerization of a single output strand is enough to take the system away
 142 from the 0 state and initiate exponential amplification. This phenomenon hampers the study of
 143 spatial communication between agents because there is only a limited time during which a PFL-
 144 bearing particle can wait for a triggering signal without self-activating itself.

145 In order to stabilize the “OFF” state (corresponding to a non-productive state) we use a species-
 146 specific deactivation template (pseudo-template, pT) that catalyzes the addition of a short 3’ tail
 147 (extension) to the trigger of the associated template (Fig. 3a). Like the autocatalytic template the

148 pseudo-template is composed of a 3' input binding part, but its output part consists of only a few
 149 nucleotides (poly-dA or poly-dT). As the trigger binds to the pT, it is elongated by the polymerase.
 150 The resulting extended strand slowly melts away (regenerating the pseudo-template so that this
 151 mechanism is catalytic) and is now unable to prime further reactions. This mechanism (Fig. S5)
 152 constitutes a signal-specific sink of tunable throughput that has been used to build multistable and
 153 excitable circuits²⁴. We adapt this strategy to particle systems by co-immobilizing the autocatalytic
 154 template with the corresponding pseudo-template on the **particles**.



155
 156 **Figure 3. Construction of 2-state agents using a bistable molecular program.** **a**, Bistable design based on a
 157 fast but saturable deactivation pathway targeting the autocatalytic species α . The two **encoding** templates
 158 ($\alpha to \alpha$ and $pT\alpha$) are co-grafted on beads to obtain bistable agents ($P\alpha_B$). **b**, Experimental setup: real-time signal
 159 is recorded either by fluorescence microscopy within 2D chambers or in test tubes (beads pellet) using a
 160 thermocycler. **c**, Panel of snapshots extracted from time-lapse fluorescence recording of individual particles $P\alpha_B$
 161 incubated with various initial amounts of α strand ($0 \leq [\alpha] \leq 40$ nM). The rightmost image shows the
 162 reconstructed kymogram. **d**, Fluorescence time traces of test tubes containing ≈ 800 $P\alpha_B$ in 10 μ L, triggered
 163 with various concentrations of strand α .

164 Microscopy (Fig. 3c) and bulk (Fig. 3d) experiments show that co-grafted particles now possess two
 165 different stable states (**nonproductive and productive**, referred as the “OFF” and “ON” states) which
 166 are selected depending on the initial conditions (respectively high or low trigger). Pseudo-templates
 167 are therefore able to abolish self-start, even in the context of the very high local concentration of
 168 templates (~ 250 μ M in the bulk of the sphere). If no signal is present in their environment, the
 169 particles stay in their **OFF** state so that the activation of bistable beads necessarily relies on the
 170 application of a stimulus exceeding a given threshold. In the following, we call the particles carrying
 171 only the **autocatalytic** template $P\alpha_M$ and the co-grafted particles $P\alpha_B$ (M and B stand for monostable
 172 and bistable, respectively).

173

174 Monostable and bistable particle encoded with a positive feedback loop

175 The dynamics of PEN DNA systems, relies on a generalist enzymatic machinery that drives the
 176 exchange of DNA signals between the rule templates: a DNA-polymerase elongates an input strand

177 that hybridizes on the input (3') side of a matching template; a nickase recognizes and cuts the
178 resulting full duplex, releasing both the input and a new output DNA. The exonuclease non-
179 specifically degrades all unprotected oligonucleotides (templates are protected), maintaining the
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181 reactions collaborate to generate a set of basic modules that can be cascaded in dynamic circuits
182 whose parameters are linked to kinetic, thermodynamic and concentration values. This versatile
183 system has already allowed the construction of various solution-phase circuits such as oscillators⁵ or
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193 fluoresces upon binding the active templates producing the α strand.** Time lapse fluorescence
194 imaging revealed the expected behavior for the PFL circuit: an initial exponential amplification phase
195 is followed by a steady-state plateau (production balances diffusion and degradation), then a return
196 to the baseline after exhaustion of dNTPs (Fig. Sx and Supplementary Movie M1). This behavior
197 closely parallels the one observed for the case of freely diffusing templates (Fig. Sx). In the case of
198 particle-tethered templates however, change in fluorescence signal is observed only on the **particle
199 position** and not in the bulk of the solution. **Therefore, the production of the output α is now
200 localized while its degradation by the exonuclease is likely to occur in the whole chamber. To assess
201 the concentration profile of α , a few autocatalytic particles are displayed in a chamber together with
202 particles functionalized by the template $\alpha\text{to}\beta$, acting like a sensor of the α strand: upon reversible
203 binding of the input strand, these particles produce an inert β strand. The reaction is monitored
204 using EvaGreen as reporter of the overall activity of each particle.****

205 **Diffusion cone + theory...**

206

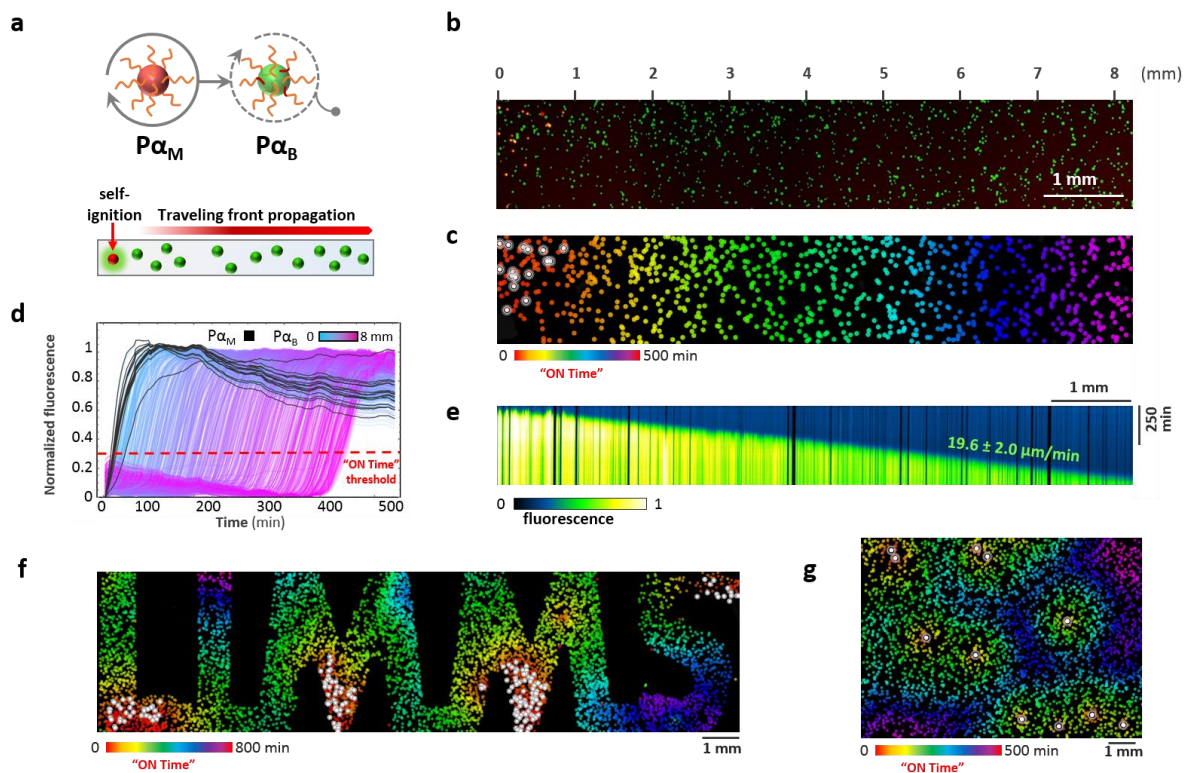
207 Notwithstanding the absence of initial trigger, particles carrying the template $\alpha\text{to}\alpha$ inevitably switch
208 to the "ON" state after a short period of incubation (Fig. XX and Fig. 2b). This phenomenon is again
209 consistent with the behavior reported for solution-phase amplification systems: autocatalytic
210 templates self-start after some time **because the first-order PFL network enforced by the dual repeat
211 template is intrinsically monostable.**²³ In order to stabilize the "OFF" state (corresponding to a non-
212 productive state) we use a species-specific deactivation template (pseudo-template, pT) that
213 catalyzes the addition of a short 3' tail (extension) to the trigger of the associated template (Fig. 2c).
214 Like the autocatalytic template the pseudo-template is composed of a 3' input binding part, but its
215 output part consists of only a few nucleotides (poly-dA or poly-dT). As the trigger binds to the pT, it
216 is elongated by the polymerase. The resulting extended strand slowly melts away (regenerating the
217 pseudo-template so that this mechanism is catalytic) and is now unable to prime further reactions.
218 This mechanism (Fig. S5) constitutes a signal-specific sink of tunable throughput that has been used
219 to build multistable and excitable circuits²⁴. We adapt this strategy to particle systems by co-
220 immobilizing the autocatalytic template with the corresponding pseudo-template on the **particles.**
221 Microscopy (Fig. XX) experiments show that co-grafted particles now possess two different stable
222 states (**nonproductive and productive**, referred as the "OFF" and "ON" states) which are selected
223 depending on the initial conditions (respectively high or low trigger **concentration**). Pseudo-
224 templates are therefore able to abolish self-start, even in the context of the very high local
225 concentration of templates ($\sim 250 \mu\text{M}$ in the bulk of the sphere). If no signal is present in their

226 environment, the particles stay in their **OFF** state so that the activation of bistable beads necessarily
 227 relies on the application of a stimulus exceeding a given threshold. In the following, we call the
 228 particles carrying only the **autocatalytic** template $P\alpha_M$ and the co-grafted particles $P\alpha_B$ (M and B
 229 stand for monostable and bistable, respectively).

230

231 **Communication: travelling front propagation in a population of bistable agents**

232 We then moved on to evaluate the communication capabilities between these **independent** agents.
 233 Figure 4a depicts an experiment where bistable particles $P\alpha_B$, initially in their OFF state, are
 234 randomly positioned in an extended channel roughly one centimeter long. On one side of the
 235 channel, dye-barcoded monostable particles $P\alpha_M$ (without pseudo-template) are introduced (Fig. 4b).
 236 As expected, $P\alpha_M$ particles **self-activate** after a short delay. The concentration inhomogeneity that
 237 they create is strong enough to trigger the switching of neighboring $P\alpha_B$, which in turn lights up and
 238 propagates the signal to other bistable particles (Fig. 4c-d and Supplementary Movie M2). This
 239 generates a travelling front that gradually converts all particles in the channel. Switch-on times for
 240 $P\alpha_B$ particles correlate linearly to the distance from the monostable triggering beads (Fig. 4e).



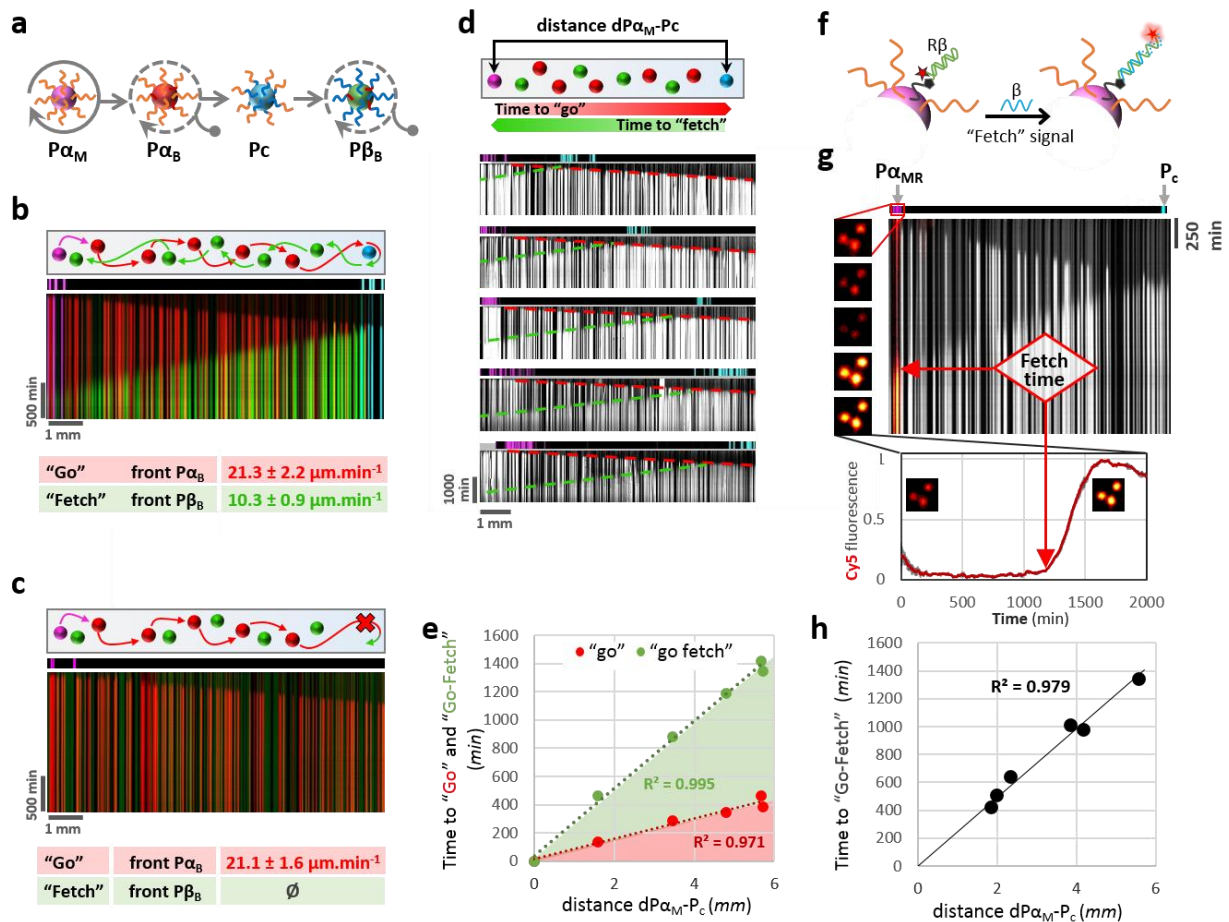
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242 **Figure 4. Travelling front propagation across a population of bistable agents ($P\alpha_B$).** *a*, Schematic
 243 representation showing the propagation of an activation signal triggered initially by the self-ignition of
 244 monostable agents ($P\alpha_M$). *b*, Barcode image (red spots = $P\alpha_M$; green dots = $P\alpha_B$). *c*, Colorized representation of
 245 the front propagation where color represents the time of amplification ("ON" time) for each particle. *The top*
 246 *bar indicates the longitudinal*~~The white dots indicate~~ *the positions of the $P\alpha_M$ (red stripes).* *d*, Fluorescence time
 247 traces extracted for each particle show the spontaneous switching of monostable particles and the correlation
 248 between the position of bistable particles and their amplification start time. *e*, The kymogram reveals the
 249 constant velocity of the travelling front across the population of $P\alpha_B$. *f, g*, Travelling front propagation across
 250 population of $P\alpha_B$ in various geometric environments. For details on image processing see Fig. [S10](#).

251

252 It is noteworthy that the front velocity is much lower than it would be if the templates and pseudo-
 253 templates were free and homogeneously distributed in the solution (Fig. S6). Indeed, production
 254 occurs only at the particle's position, on non-diffusing templates, while the degradation of diffusing
 255 signal happens in the whole chamber (thus limiting the amount of triggers able to diffuse from bead
 256 to bead). The velocity of the front propagation can be finely tuned between a few to tens of
 257 micrometers per minute by tinkering with the agents' features (ratio template/pseudo-template, Fig.
 258 S7) or the experimental conditions (such as enzymes concentration or particle density, Fig. S8-9).
 259 Additionally, these dynamic activation patterns have been generated in various geometric
 260 environments (Fig XX).

261



262

263 **Figure 5. "Go-Fetch" program using 4 agent populations.** **a**, Schematic representation showing the flow of
 264 information between the four bead types. **b**, Kymogram representation of the experiment. Evagreen
 265 fluorescence signals associated with the beads are color-coded according to the bead type (magenta = $P\alpha_M$, red
 266 = $P\alpha_B$, cyan = $P\alpha_{\beta}$, green = $P\beta_B$). **c**, Control experiment with no converter bead at the end of the channel. **d**,
 267 Converter beads are positioned at different locations in the channel. **e**, Front travelling time as a function of
 268 sender-target distance in **d**. **f**, An additional mechanism allows the sender bead to detect the arrival of the
 269 return front. **g**, Experimental snapshots, kymogram and plot of the red fluorescence for sender particles. **h**,
 270 Light-up time of the sender particle as a function of its distance to the nearest converter.

271 Programmability: a 4-agent network

272 Communication between the particles need not be limited to a single molecular pathway: as
 273 observed in bacterial quorum sensing, various agents can possess distinct circuits, producing and
 274 detecting orthogonal signaling molecules²⁵. To demonstrate this concept, we designed a network
 275 using two signaling compounds and four different kinds of agents. First, we created a second type of

276 bistable particles producing and emitting strand β (Fig. S11). We confirmed the orthogonality of the
277 two pathways in travelling front experiments using a mixed population of bistable agents P_{α_B} and
278 P_{β_B} . α and β fronts were set to propagate in opposite directions, and affected only the
279 corresponding particles (Fig. S12). In a second step, the initiation of the front propagating through
280 the P_{β_B} population was coupled to the arrival of the front producing α (Fig. 5a), **using converter**
281 **particles $P_{\alpha \rightarrow \beta}$, carrying the template α to β (which take α as input to produce β as output)**. The
282 topography of the spatial network, nicknamed “Go-Fetch”, is as follows: bistable agents P_{α_B} and P_{β_B}
283 are mixed together in a channel. At one extremity, a few monostable P_{α_M} (the sender particles) are
284 deposited; on the opposite side of the channel, the converter agents $P_{\alpha \rightarrow \beta}$ represent the target.
285 Figure 5b (see also Fig. S13 and Supplementary Movie M3) shows sequentially i) the **self-activation**
286 of the monostable beads P_{α_M} , ii) the propagation of the signal through the bistable population P_{α_B} ,
287 iii) the activation of the converter particles (which light on once the front reaches their position) and
288 iv) the initiation and propagation of a second front travelling back to the initial sender, through the
289 bistable particles P_{β_B} .

290 **Satisfyingly,** A control experiment without converter agents also produced a first front propagation
291 through particles P_{α_B} , but the return front was not generated and all particles P_{β_B} stayed “OFF” for
292 the duration of the experiment (30 hours). These results demonstrate the use of multiple speciated
293 agents collaborating in a unique, spatially and time-resolved task. This program can for example be
294 used by a sender particle to infer its distance to the nearest target particle, over a scale that is 3
295 orders of magnitude larger than the agents themselves. By arbitrarily varying the position of the
296 $P_{\alpha \rightarrow \beta}$ in the channel, we indeed observed that the sender particle receives this distance information
297 as the time needed for the signal to return, with a linear relationship between distance and time (Fig.
298 5d-e). It is possible to imagine that the particles use this “**chemical sonar**” to control downstream
299 processes **...** As a basic model of differentiation, we added a module to the sender particles so that
300 they light up as soon as the system has fetched the distance information (Fig. 5f): P_{α_M} are modified
301 with a molecular beacon-like probe, which fluoresces upon binding and polymerization of the
302 returning β strands (Fig. 5g). The full system now differentiates the sender particles P_{α_M} with a
303 timing that is precisely controlled by their **position** relative to the nearest converter particles $P_{\alpha \rightarrow \beta}$
304 (Fig. 5h)¹⁵.

305 One may note that the return front is much slower than the first front passing through P_{α_B} . It is also
306 twice slower than the propagation through P_{β_B} initiated by P_{β_M} (Fig. S13), demonstrating that this is
307 not due to the P_{β_B} propagation system itself being intrinsically slower. We therefore conclude that it
308 is the presence of *active* P_{α_B} that slows down the propagation of the β front. This observation
309 suggests that competition for catalytic resources, such as nickase and/or polymerase, happens even
310 if the two networks are spatially separated and occupy only a minor fraction of the total enzymatic
311 mixture^{26,27} (see also Fig. S9). **Competition/depletion/concentration enzymes ???**

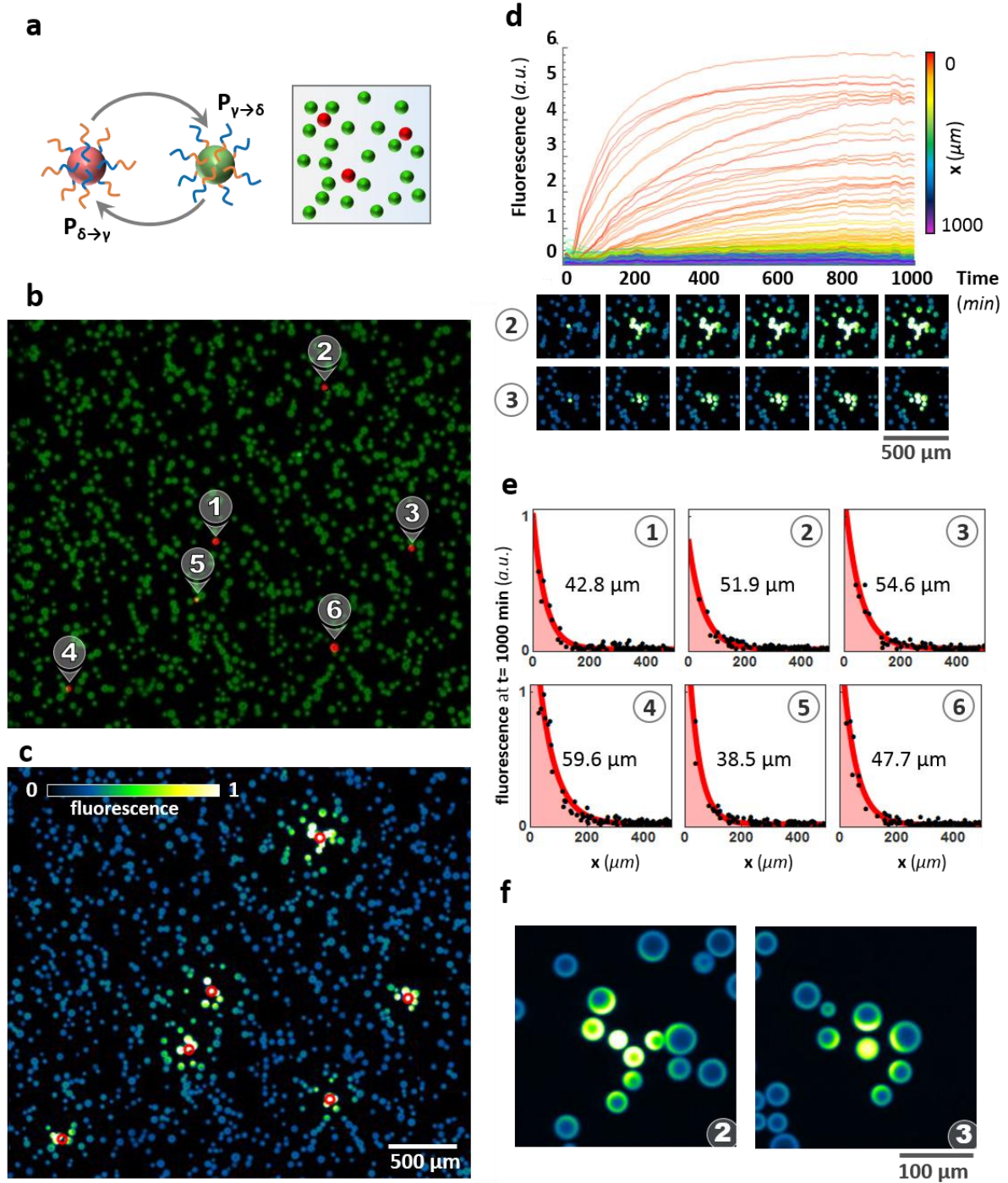
312 **Colony formation by symbiotic particles**

313 While the previous experiments show that beads can behave as **independent** entities, it is also
314 possible to engineer synergic dependency and cooperativity among the individual agents. To
315 evaluate this possibility, we split an autocatalytic loop into two templates (one encoding γ to δ and
316 one δ to γ), which are separately attached to different microspheres (Fig. Xa). An active PFL network
317 now necessarily requires oligonucleotides to be exchanged between both bead types. We created
318 flat **fluidic** chambers containing a high density of $P_{\gamma \rightarrow \delta}$ particles (10^5 part./cm²) and **a few** $P_{\delta \rightarrow \gamma}$
319 particles (Fig. XXb). Real-time fluorescence monitoring revealed the emergence of localized
320 activation colonies located around the minority particles (Figure 6Xc, S15 and Supplementary Movie
321 M4). These **colonies** emerge and grow to reach a steady state (Figure 6Xd). The activity within the
322 majority population is directly related to the presence of a minority bead in their neighborhood. .
323 The steady state fluorescence of the majority beads decays quickly with the distance to their feeding
324 $P_{\delta \rightarrow \gamma}$, and fitting yield a characteristic length on the order of 50 μ m (Fig. XXe, see Supplementary

325 Note 3 for justification of the exponential decay model). Interestingly, we observe anisotropy in the
326 particle reactivity, the side of the $P_{\gamma \rightarrow \delta}$ exposed to the $P_{\delta \rightarrow \gamma}$ being more active (Fig. Xd) at steady
327 state ?. This confirms that the system dissipate enough free energy to maintain strong steady-state
328 gradients over micrometric ($\sim 10 \mu\text{m}$) distances. (in the absence of a active sustaining mechanism
329 such gradient would equilibrate in $r^2/D = \text{xx minute}$)
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332 In the experiment depicted in Figure XX, we gradually increased the density of minority particles
333 from XX to XX part./cm². At low $P_{\delta \rightarrow \gamma}$ densities (XX to XX part.cm²), the colonies are small, well
334 defined, and similar for all conditions. However, when the density of minority beads reaches a
335 threshold, we observe a sudden increase in colony size and much more activity in the population of
336 majority beads. Since this happens when the mean distance between minority beads becomes close
337 to the colonies characteristic length, it suggests that a colony-to-colony cooperation activation
338 mechanism comes to play, where the signal strand diffusing from one colony is able to activate
339 majority beads from a second colony, reinforcing it, and benefiting in return. We therefore
340 observe a ~~double~~ two-tier communication mechanism where $P_{\gamma \rightarrow \delta}$ beads locally detect the presence
341 of a minority bead in their vicinity, but also globally sense the average density of the minority
342 partners through a percolation mechanism associated with a sharp density threshold.
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Figure 6. Mixed populations of symbiotic particles form localized clusters of activity around the minority agents. *a*, Schematic representation of the system. *b*, Barcode image of a portion of the fluidic chamber (Green = $P_{\gamma \rightarrow \delta}$, Red = $P_{\delta \rightarrow \gamma}$). *c*, Snapshot image of EvaGreen fluorescence at $t = 1000$ min. The minority particles are indicated by red circles. *d*, The fluorescence of each majority particle is plotted as a function of its distance to the nearest minority particle. The bottom images represent fluorescence snapshots of two colonies at the corresponding time. *e*, Profile of reactivity for each colony. The fluorescence of each majority particle is plotted as a function of its distance to the minority particle and fitted with a simple exponential decay (length parameters l_d are indicated in black). *f*, Confocal image (10X magnification) of two colonies showing the anisotropy of reactivity for the surrounding majority particles.

356 ~~Figure 6. Mixed populations of symbiotic particles form localized clusters of activity around the minority~~
357 ~~agents. a, Schematic representation of the system. b, Snapshot of a portion of the fluidic chambers at t = 300~~
358 ~~min. The minority particles (indicated by red crosses) are present at increasing concentrations in the various~~
359 ~~experiments (their total number in each chamber is given in parenthesis). c, Time lapse images for the two~~
360 ~~labelled clusters in chamber #4 and #5. Images are taken every 30 minutes. d, Profile of activity for two clusters~~
361 ~~from chamber #4. The fluorescence of each majority bead is plotted as a function of its distance to the minority~~
362 ~~particle and fitted with a simple exponential decay (length parameters are 100 (top) and 63 μm (bottom)). The~~
363 ~~inset shows the corresponding cluster. e, Plot of the mean distance between two minority particles and the~~
364 ~~activity that they generate among the majority particles, for various densities of minority particles. The activity~~
365 ~~is defined as the number of majority particles which cross a threshold of fluorescence.~~

366 Conclusion

367 Communication using diffusive compounds is a general biological strategy to build multi-agents
368 behaviors at microscopic scales. Diffusive cell signaling is typically classified into autocrine (self-
369 communication) or paracrine signaling (cross-communication)²⁸. These two types are naturally
370 implemented in our system because the encoding templates can be designed to use either
371 endogenous, exogenous or both signals as inputs.

372 To better understand the scaling associated with the usage of diffusive communication between
373 agents, we analytically solved the concentration profile generated by the shell-like reacting particles
374 (Supplementary Note 4). The mathematical expression depends on the dimensionality of the
375 problem²⁹, but the length parameter is always controlled by $\sqrt{D/k}$, where D is the diffusion rate and
376 k the first-order degradation rate for the signaling compound. In our conditions, these parameters
377 are $D = 18.10^3 \mu\text{m}^2 \cdot \text{min}^{-1}$ and $k = 0.5 \text{ min}^{-1}$, which predicts a length scale of 200 μm , in agreement
378 with our observations (Fig. 5d and Supplementary Note 3). A particle thus communicates efficiently
379 with a neighborhood that is one order of magnitude larger than its size, enabling the unfolding of
380 collective behaviors over relatively large distances.

381 Our strategy to give memory and differentiated states to the agents involves autocrine bistable
382 circuits based on PFL^{30,31}. In the absence of physical boundaries, this approach requires that each
383 particle is able to **autonomously** maintain its high state, i.e. compensate for the signals lost by
384 diffusion. Solving the previous source and decay model in the case of an isolated particle carrying a
385 positive feedback loop exposes the experimental requirements (Supplementary Note 4 and 5): our
386 model predicts the existence of a critical size, or critical grafting density, below which a particle loses
387 the ability to maintain a differentiated state. In other words, competition between global and local
388 effects imposes that smaller agents dissipate faster in order to maintain their individuality.

389 Such competition between global and local effects has already been pointed out for other tethered
390 DNA systems³² and imposes constraints on the miniaturization of molecular programming
391 approaches. Compartmentalization techniques, such as microfabricated chambers, vesicles or
392 emulsions³³ provide an alternative to direct tethering of the DNA rules. Compartments efficiently
393 restrict the diffusion of reactive species³⁴, but conversely require specific strategies in order to open
394 communication channels between the units³⁵⁻³⁸. Engineered microorganisms have also been used to
395 implement collective behaviors, although the programming of living systems for such tasks remains
396 challenging³⁹⁻⁴³.

397 **Our approach based on template-decorated particles provides the basic set of tools required to build**
398 **and study artificial microscopic communities¹⁹. It uses a hierarchical approach that maintains a**
399 **rational link from molecular pairwise interactions all the way to multi-particles dynamics. Compared**
400 **to previous work in the study of collective behaviors⁴⁵⁻⁴⁷, the main advantage is a high level of**
401 **programmability⁴⁴ i.e. that fact that one can freely decide the set of signals to which a given particle**
402 **is sensitive, and its precise chemical behavior in response to these signals. Our example with the go-**
403 **fetch network has shown that, like in a multicellular organism, particles carrying different sets of**
404 **templates can take on the various specialized roles necessary to achieve a global task. The colony**

405 *experiment demonstrates fine sensing of the features of the local and global environment. While the*
406 *particles used here are too large to display Brownian motion, one may envision that microscopic*
407 *agents whose motility is controlled by the environment-sensitive network they carry could provide a*
408 *path to dynamically self-shaping materials. With a straightforward synthesis, they open the route to*
409 *a massive scale-up in the complexity of man-made molecular computing systems. In particular, they*
410 *will bring a high level of programmability to the study of collective behaviors, and the modelling of*
411 *biological morphogenesis.* In practical diagnostic applications, we also envision that each particle
412 could be programmed to **independently** perform a different sensing task, while collaborating with
413 the others to provide a global consensus decision⁴.

414 **Methods**

415 Methods and associated references are available in the **Supplementary files**.

416 **Acknowledgments**

417 This work was supported by the JSPS with a Grant-in-Aid from the JSPS for Scientific Research on
418 Innovative Areas “Synthetic Biology for Comprehension of Biomolecular Networks” (#23119001),
419 and an ERC Consolidator grant “ProFF” (#647275). G.G. acknowledges financial support by the JSPS
420 Postdoc program. We thank Nicolas Bredeche and Nathanael Aubert-Kato and Anthony Genot for
421 advices and Yannick Tauran and Alexandre Baccouche for expressing and purifying the exonuclease.
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423 **Author contributions**

424 G.G. designed the study, performed experiments, analyzed the data and wrote the manuscript. A.Z.
425 did mathematical analysis, contributed to image analysis and manuscript writing. J.C.G, A.E.T.
426 contributed to experimental setup and designed the study. T.F. provided support with the
427 microfluidic platform. Y.R. conceived, designed, and supervised the study, analyzed the data and
428 wrote the manuscript.

429 **Additional information**

430 Supplementary information is available for this paper. Reprints and permissions information is
431 available online at Correspondence and requests for materials should be addressed to Y.R.

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